

# Genetic Diversity of White-spotted Rabbitfish (*Siganus canaliculatus*) on Different Seagrass Habitats in Inner Ambon Bay, Indonesia Based on Mitochondrial CO1 Sequences

*by* Husain Latuconsina

---

**Submission date:** 18-Apr-2024 09:24AM (UTC+0700)

**Submission ID:** 2353397912

**File name:** 10\_Latuconsina\_et\_al.\_2024\_TLSR.pdf (1.91M)

**Word count:** 8388

**Character count:** 44975



3  
**Genetic Diversity of White-spotted Rabbitfish (*Siganus canaliculatus*) on Different Seagrass Habitats in Inner Ambon Bay, Indonesia Based on Mitochondrial CO1 Sequences**

**Authors:**

Husain Latuconsina,\* Nurlisa A. Butet, Ridwan Affandi, M. Mukhlis Kamal, Syamsul Bachry and Agus A. Hakim

\***Correspondence:** husain.latuconsina@unisma.ac.id

**Submitted:** 11 November 2022; **Accepted:** 2 May 2023; **Published:** 2 April 2024

**To cite this article:** Husain Latuconsina, Nurlisa A. Butet, Ridwan Affandi, M. Mukhlis Kamal, Syamsul Bachry and Agus A. Hakim. (2024). Genetic diversity of white-spotted rabbitfish (*Siganus canaliculatus*) on different seagrass habitats in inner Ambon Bay, Indonesia based on mitochondrial CO1 sequences. *Tropical Life Sciences Research* 35(1): 273–291. <https://doi.org/10.21315/tlsr2024.35.1.15>

**To link to this article:** <https://doi.org/10.21315/tlsr2024.35.1.15>

**Highlights**

- 2
- The genetic diversity (haplotype and nucleotide variability) of white-spotted rabbitfish (*Siganus canaliculatus*) population living in different seagrass habitats in Inner Ambon Bay (IAB) waters.
  - Intra- and inter-population genetic differentiation of *Siganus canaliculatus* were 21.19% dan 78.81%, respectively. Intra- and inter-population genetic distances were in range of 0.40–1.13 and 0.00–0.37.
  - The pattern and direction of tidal currents as a link or barrier to spatial distribution and connectivity of *S. canaliculatus* larvae between seagrass habitats, as well as the presence of different anthropogenic pressures in each seagrass habitat, are thought to influence the genetic characteristics (genetic diversity, genetic variation, genetic differentiation, and genetic distance) of *S. canaliculatus* populations in IAB waters.

**TLSR, 35(1), 2024**

© Penerbit Universiti Sains Malaysia, 2024

## Genetic Diversity of White-spotted Rabbitfish (*Siganus canaliculatus*) on Different Seagrass Habitats in Inner Ambon Bay, Indonesia Based on Mitochondrial CO1 Sequences

<sup>1</sup>Husain Latuconsina,<sup>\*</sup> <sup>2</sup>Nurlisa A. Butet, <sup>2</sup>Ridwan Affandi, <sup>2</sup>M. Mukhlis Kamal, <sup>3</sup>Syamsul Bachry and <sup>2</sup>Agus Alim Hakim

<sup>1</sup>Department of Biology, Faculty of Mathematics and Natural Sciences, Universitas Islam Malang, Dinoyo, Malang 65144, Indonesia

<sup>2</sup>Department of Aquatic Resources Management, Faculty of Fisheries and Marine Science, IPB University, FPIK Bld. 3rd Floor, Kampus IPB Dramaga, Bogor 16680, Indonesia

<sup>3</sup>Program Study of Biology, Universitas Pahlawan Tuanku Tambusai, Jl. Tuanku Tambusai No. 23 Bangkinang Kota, Kampar, Riau 28412, Indonesia

**Submitted:** 11 November 2022; **Accepted:** 2 May 2023; **Published:** 2 April 2024

**To cite this article:** Husain Latuconsina, Nurlisa A. Butet, Ridwan Affandi, M. Mukhlis Kamal, Syamsul Bachry and Agus A. Hakim. (2024). Genetic diversity of white-spotted rabbitfish (*Siganus canaliculatus*) on different seagrass habitats in inner Ambon Bay, Indonesia based on mitochondrial CO1 sequences. *Tropical Life Sciences Research* 35(1): 273–291. <https://doi.org/10.21315/tlsr2024.35.1.15>

**To link to this article:** <https://doi.org/10.21315/tlsr2024.35.1.15>

**Abstract:** This study aims to analyse the genetic diversity of *Siganus canaliculatus* in the Inner Ambon Bay (IAB) waters. DNA of *S. canaliculatus* specimens collected from IAB was extracted from tissues using a Tissue Genomic DNA Mini Kit, and partial CO1 genes were amplified using pair of universal primers. Genetic distances were determined by Kimura 2-parameter, and phylogenetic trees were constructed using the neighbour-joining method in MEGA 10.2.2 software. Arlequin software was used to analyse Fixation Index (Fst) and Analysis of Molecular Variance (AMOVA). There are three SNPs of *S. canaliculatus* from IAB that distinguish GenBank sequence data from *S. canaliculatus*. In Tanjung Tiram population group, contained three specific 677 (A), 679 (G), 703 (T) sites and two 693 (G), 714 (A) sites for the Nania population. Haplotype and nucleotide diversity of each population range from 0.000 to 1,000 and 0.000 to 0.004. Intra- and inter-population genetic differentiation were 21.19% dan 78.81%, respectively. Intra- and inter-population genetic distances were in range of 0.40–1.13 and 0.00–0.37, respectively. The pattern and direction of tidal currents as a link or barrier to spatial distribution and connectivity of *S. canaliculatus* larvae between seagrass habitats, as well as the presence of different anthropogenic pressures in each seagrass habitat, are thought to influence the genetic characteristics (genetic diversity, genetic variation, genetic differentiation and genetic distance) of *S. canaliculatus* populations in IAB waters. The results of this study provide information about the urgency of habitat-based fisheries management to support sustainable utilisation.

**Keywords:** Genetic Differentiation, Phylogenetic, Genetic Distance, Genetic Diversity, Single Nucleotide Polymorphisms (SNPs).

<sup>\*</sup>Corresponding author: [husain.latuconsina@unisma.ac.id](mailto:husain.latuconsina@unisma.ac.id)

## INTRODUCTION

The white-spotted rabbitfish (*Siganus canaliculatus* Park, 1797) is a teleosts fish belonging to the family Siganidae and order Perciformes (Woodland 2001). This rabbitfish is widely distributed in the Indo-Pacific region by utilising tidal rhythms in three important habitats, namely mangroves, seagrass beds and coral reefs (Allen & Erdmann 2012; Latuconsina *et al.* 2013; Latuconsina *et al.* 2023). In the Inner Ambon Bay (IAB hereafter), *S. canaliculatus* occurs in a variety of seagrass habitats with different densities and diversity of vegetation (Ambo-Rappe *et al.* 2013; Latuconsina *et al.* 2020a), enough to influence and determine the variation of growth and reproduction biology (Latuconsina *et al.* 2022).

The IAB waters are naturally separated from Outer Ambon Bay (OAB) by a narrow inlet (sill) with a depth of  $\pm 12$  m, causing the water mass circulation not to run smoothly (Basit *et al.* 2012; Noya *et al.* 2016a; Saputra & Lekalette 2016; Salamena *et al.* 2021). Various anthropogenic activities threaten the existence of seagrass habitats in IAB, such as garbage disposal, organic waste and dredging of beach sand (Evans *et al.* 1995; Selano *et al.* 2009; Manullang *et al.* 2021), sedimentation (Irawan & Nganro 2016; Noya *et al.* 2016b; Rahmawan *et al.* 2019). The condition of the waters of IAB which is more influenced by anthropogenic nutrient inputs from the mainland through river flow, and the supply of nutrients from the OAB from the influence of upwelling in the Banda Sea which can increase primary productivity and trigger eutrophication in IAB waters (Basit *et al.* 2012; Pello *et al.* 2014, Mahmudi *et al.* 2020); Contamination of heavy metals, Cadmium (Cd) and Chromium (Cr) in sediments, nitrate and phosphate content is already high and in the nutrient contaminated category due to the influence of runoff from land and input from rivers (Ikhsani *et al.* 2016). Human settlements, land use around IAB, and semi-enclosed morphological conditions of IAB, as well as the number of river estuaries are factors that trigger the decline in quality of IAB waters (Gemilang *et al.* 2017; Salamena *et al.* 2021).

High ecological pressure in IAB waters affects the population dynamics of *S. canaliculatus*. Several authors (Manik 1998; Latuconsina *et al.* 2020b) found that the higher fishing mortality and exploitation rate of *S. canaliculatus* without considering the size catch limit resulted in fishing effort are economically unfeasible. Moreover, overexploitation and habitat degradation has caused a decrease in genetic diversity (Toro & Caballero 2005; Markert *et al.* 2010; Martinez *et al.* 2018). Recently, Madduppa *et al.* (2019) reported the occurrence in genetic diversity of the rabbitfish comparing the north and south of Jakarta Bay population in which the environmental influences and the impact of anthropogenic activities showed a greater impact.

This research is a preliminary study using mitochondrial CO1 sequences, to determine whether differences in the physical characteristics of seagrass habitat and geographical distance in IAB can affect the genetic diversity and genetic distance of *S. canaliculatus*. The research results are expected to become scientific information to support habitat-based fisheries management.

## MATERIALS AND METHODS

### Collection Sample

A total of 14 individual *S. canaliculatus* (samples) were collected from fish sampling locations in different seagrass habitats. The first habitat is the seagrass mixed vegetation located at two observation stations; Station 1: Tanjung Tiram with as many as three samples, and Station 2: Halong with as many as four samples. Meanwhile, in the monospecific seagrass meadows, there are two observation stations which are both drained by rivers, namely; Station 3: Poka with as many as three samples, and Station 4: Nania with as many as four samples (Fig. 1).



**Figure 1:** Map of research stations in Inner Ambon Bay Waters (IAB).

Mixed vegetation seagrass habitat (five species of seagrass: *Enhalus acoroides*, *Thalassia hemprichii*, *Halophila ovalis*, *Cymodocea rotundata* and *Halodule pinifolia*) at two observation stations. Station 1: Tanjung Tiram ( $3^{\circ}39'16.5''\text{S}$  and  $128^{\circ}12'0.43''\text{E}$ ) dominated by fine sand substrate, around which mangrove vegetation grows, but not dense because it is very close to residential areas. Station 2: Halong ( $3^{\circ}39'32.9''\text{S}$  and  $128^{\circ}12'31.2''\text{E}$ ) is dominated by coarse sand substrate, around which there is a coral reef rehabilitation/transplant area. Whereas monospecific seagrass meadows (only one species of seagrass, *Enhalus acoroides*) includes two observation stations which both have river mouths. Station 3: Poka ( $3^{\circ}38'36.48''\text{S}$  and  $128^{\circ}11'42.54''\text{E}$ ) with a sandy mud substrate, and far from the coastline and has a rehabilitation mangrove area although not extensive. Station 4: Nania ( $3^{\circ}37'58.7''\text{S}$  and  $128^{\circ}13'45.1''\text{E}$ ) dominated by sandy mud substrate, around which natural mangrove vegetation grows which is quite dense, is located far from the coastline, and has been designated as a conservation area by the government local (Latuconsina *et al.* 2020a).

### **Isolation and purification**

Tissue samples from 14 fish were preserved in absolute ethanol. Genomic DNA extraction using the Mini Kit for Tissue Genomic DNA Mini Kit (Gene Aid) Cat. No. GT050 (Vogelstein & Gillespie 1979) according to the Spin-Column protocol with modifications.

### **Amplification and sequencing**

The nucleotides of the COI gene were amplified using polymerase chain reaction (PCR). The COI primer used is a universal aquatic biota primer with the primary name COI-F (5'-GGTCAACAAATCATAAAATATTGG-3') and primary COI-R (5'-TAAACTTCAGGGTGACCAAAAAATCA-3') with a sequence length of 700 bp (Butet 2003, unpublished). Amplification uses the Labcycler Squence engine.

The volume of the mixed reaction composition is 25 µL with 4.5 µL ddH<sub>2</sub>O, 12.5 µL My Tag HS Red Mix, 1.5 µL primer forward and 1.5 µL reverse primer and 5 µL template DNA. Temperature PCR is pre denaturation 94°C (3 min), followed by denaturation for 35 cycles at 94°C (45 s), annealing 54°C (1 min), elongation 72°C (30 s), and final extension 72°C (5 min). Further sequencing was carried out in the First Base laboratory (Malaysia), using amplified DNA stored in agarose gel 1.2% (Sambrook et al. 1989).

### **Data Analysis**

Nucleide editing and alignment uses Clustal W in MEGA 10.2.2 program (Kumar et al. 2018). Analysis of genetic diversity which includes haplotype diversity (Hd) and nucleotide diversity ( $\pi$ ) uses the DnaSP program version 510.01 (Rozas 2009) and Network version 5 (Bandelt et al. 1999). Analysis Molecular Variance (AMOVA) and Fixation Index (Fst) were analysed through the Arlequin program (Excoffier et al. 1992). Intra and interpopulation genetic distance (D) and phylogenetic tree based on the neighbour-joining (NJ) method with the Kimura 2-parameter (K2P) model with bootstrap 1000 repetition uses MEGA 10.2.2 programme (Kumar et al. 2018). Sample identification based on sequence similarity approach was carried out using Barcoding of Life Database (BOLD) (<http://www.barcodinglife.org>) and GenBank (<http://www.ncbi.nlm.nih.gov>). The in-group sequences of the COI gene from GenBank consist of *S. canaliculatus* (KJ872545) from China, *S. fuscescens* (EF025185) from China, *S. sutor* (MG677546) from South Korea, and out-group *Lutjanus russellii* (EF514208) from China.

## RESULTS

### Genetic Diversity of White-Spotted Rabbitfish

A total of 14 sequence samples that were BLASTn at NCBI and BOLD Systems showed an average identity of the four populations, representing observation stations from IAB waters (Tanjung Tiram, Halong, Poka and Nania) varied between 98.37% to 98.96% with a query cover of 100% with the species *S. canaliculatus* (KJ872545.1) origin in China.

### Single Nucleotide Polymorphism

Table 1 shows the comparison between COI gene marker sequences results of *S. canaliculatus* found in IAB waters and KJ872545.1 from China. At 700 bp, it was obtained three sites of single nucleotide polymorphisms (SNPs) namely sites 16 (A), 229 (A) and 379 (C).

**Table 1:** Single nucleotide polymorphism of the *S. canaliculatus* from IAB and China.

Species/ Access code	Nucleotide site															
	1	2	2	79	0	1	1	4	7	7	8	9	0	0	1	1
	6	0	9	9	5	4	9	1	7	9	4	3	1	3	3	4
<i>S. canaliculatus</i> (KJ872545.1)*	G	C	G	T	C	T	A	A	G	T	G	A	T	C	G	G
<i>S. canaliculatus</i> Tanjung Tiram-1	6 A	T	A	C	T	C	.	.	A	G	A	.	C	T	.	.
<i>S. canaliculatus</i> Tanjung Tiram-2	A	T	A	C	T	C	.	.	A	G	A	.	C	T	.	.
<i>S. canaliculatus</i> Tanjung Tiram-3	A	T	A	C	T	C	.	.	A	G	A	.	C	T	.	.
<i>S. canaliculatus</i> Halong-1	A	.	A	C	.	.	.	.	.	.	.	.	.	.	A	.
<i>S. canaliculatus</i> Halong-2	A	.	A	C	T	C	.	.	.	.	A	.	.	.	A	.
<i>S. canaliculatus</i> Halong-3	A	.	A	C	T	C	.	.	.	.	A	.	.	.	.	.
<i>S. canaliculatus</i> Halong-4	A	.	A	C	.	.	.	.	.	.	.	.	.	.	.	.
<i>S. canaliculatus</i> Poka-1	A	T	A	C	T	C	.	.	.	.	A	.	C	.	.	.

(continued on next page)

**Table 1:** (continued)

Species/ Access code	Nucleotide site															
	1	2	2	79	0	1	1	4	7	7	8	9	0	0	1	1
	6	0	9	9	5	4	9	1	7	9	4	3	1	3	3	4
<i>S. canaliculatus</i> Poka-2	A	T	A	C	T	C	.	.	.	.	A	.	C	.	.	.
<i>S. canaliculatus</i> Poka-3	A	T	A	C	T	C	.	.	.	.	A	.	C	.	.	.
<i>S. canaliculatus</i> Nania-1	A	.	A	C	T	C	.	G	.	.	A	<b>G</b>	C	.	A	<b>A</b>
<i>S. canaliculatus</i> Nania-2	A	.	A	C	T	C	G	.	.	.	A	<b>G</b>	C	.	A	<b>A</b>
<i>S. canaliculatus</i> Nania-3	A	.	A	C	T	C	G	G	.	.	A	<b>G</b>	C	.	A	<b>A</b>
<i>S. canaliculatus</i> Nania-4	A	.	A	C	T	C	.	.	.	.	A	<b>G</b>	C	.	A	A

Note: \* = GenBank origin species.

Based on haplotype and nucleotide variability, the genetic diversity of white-spotted rabbitfish (*S. canaliculatus*) population living in different seagrass habitats in IAB waters is presented in Table 2.

**Table 2:** Genetic diversity of *S. canaliculatus* on seagrass habitats in IAB.

Type of seagrass habitat	Research station	n	Genetic diversity		
			Hn	Hd	$\pi$
Mixed vegetation	Tanjung Tiram	3	1	0.000	0.000
	Halong	4	4	1.000	0.004
Single vegetation	Poka	3	1	0.000	0.000
	Nania	4	4	1.000	0.002

Note: n = number of samples, Hn = number of haplotypes (Hn), Hd = Haplotype diversity,  $\pi$  = nucleotide diversity

The genetic differentiation of the population between observation stations in IAB waters showed a significant difference between the Fst and AMOVA tests (0.79;  $p < 0.05$ ) (Table 3).

**Table 3:** Genetic differentiation (Fst) of Inter and intra-population on *S. canaliculatus* based on different seagrass habitats in IAB.

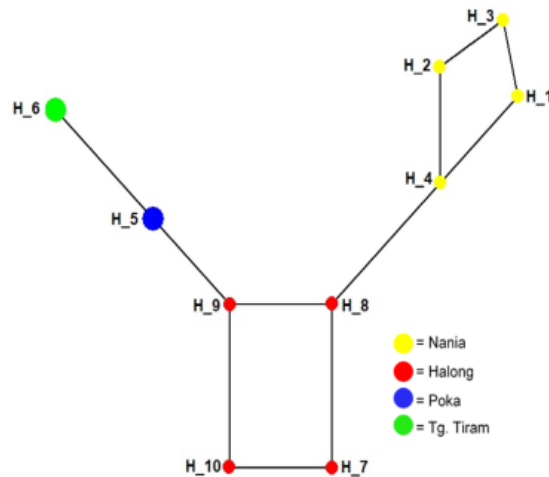
Source of variation	d.f.	Variation (%)	Fst	p-value
Inter-population	3	78.81	0.79	<0.05
Intrapopulation	10	21.19		
Total	13			

Note: d.f = degree of freedom. Fst = Genetic differentiation



### Haplotype

The results of the Network Median Joining analysis for the inter-population of *S. canaliculatus* from four seagrass habitats (Fig. 2) show that the haplotype construction forms three clusters.



**Figure 2:** Median joining network of inter-population haplotypes of *S. canaliculatus* in different seagrass habitats in IAB forming three population groups.

### Genetic Distance

Intra- and inter-population genetic distances of *S. canaliculatus* in seagrass habitats differed in IAB waters by 0.00–0.37, and 0.40–1.13, respectively. The lowest intra-population genetic distance, 0.00, was found in Tanjung Tiram and Poka, while the highest genetic distance at Halong station was 0.37 (Table 4).

**Table 4:** Genetic distances of *S. canaliculatus* CO1 gene based on Kimura 2-parameter model.

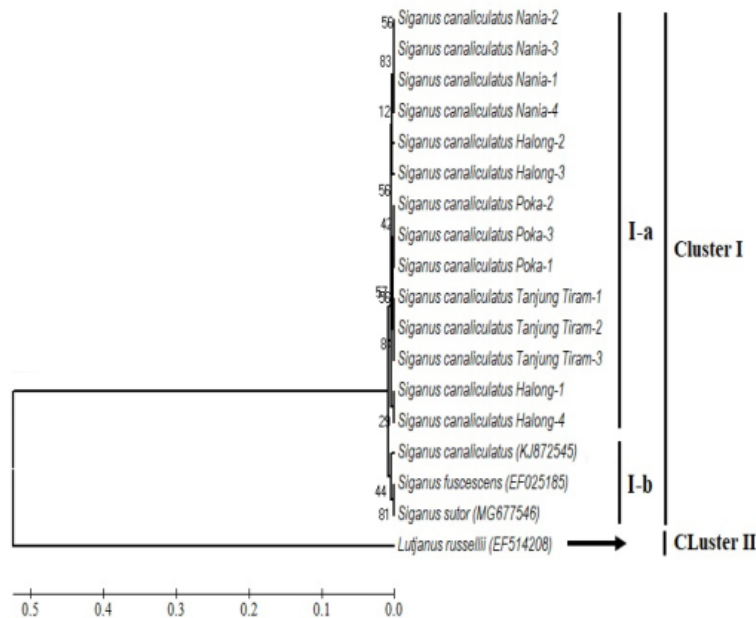
Site	Tanjung Tiram	Halong	Poka	Nania
Tanjung Tiram	0.00*			
Halong	1.00	0.37*		
Poka	0.40	0.58	0.00*	
Nania	1.13	0.86	0.73	0.17*

Note: \* = intra-population mean genetic distance of *S. canaliculatus* in IAB.

The low inter-population genetic distance, Tanjung Tiram and Poka, were 0.40. The highest was Tanjung Tiram and Nania at 1.13 (see Table 4). Population in Tanjung Tiram and Poka have the closest kinship, it is because the two populations are still in close water areas compared to the distance between the Tanjung Tiram population and the Nania population which is quite far.

### Phylogenetic

The interpopulation phylogenetic reconstruction of *S. canaliculatus* from IAB and *Siganus* spp. from other worlds is based on the NJ models with 100 bootstrap repetitions (Fig. 3). It is known that the population of *S. canaliculatus* is far from the population of China and South Korea with bootstrap values of 10 to 85. However, it was seen that two individuals (Halong-1 and Halong-4) were slightly separated from other individuals and clustered differently, with a genetic distance of 0.37 (0.37%) and a bootstrap value of 30.



**Figure 3:** Neighbour-joining (NJ) phylogenetic tree reconstruction using 14 sequences of *S. canaliculatus* based on Kimura 2-parameter model and 1,000 bootstrap replicates.

## DISCUSSION

CO1 gene is a gene that has effectiveness in validating species at the intraspecies and interspecies levels (Ward *et al.* 2005). Species that have a similarity level (> 97%) are the same or closely related species (Hebert *et al.* 2003). The COI gene is a genetic marker that can also provide clear information on changes in the sequence of nucleotide bases between species (Hajibabaei *et al.* 2006; Ward *et al.* 2008). Therefore, it has been widely used for animal identification. In addition, the COI gene can identify reproductively isolated groups.

The various sites are specific locations found exclusively on *S. canaliculatus*, serving as the species barcode within the IAB waters. In addition, specific sites were found at Tanjung Tiram with specific sites [677 (A), 679 (G), 703 (T)], and Nania [693 (G), 714 (A)] can be seen in Table 1, while the Halong and Poka populations do not have specific nucleotide sites. Intra- and inter-population genetic variation in different seagrass habitats is proven by the presence of nucleotide polymorphisms (specific nucleotide sites). This indicates the possibility of gene mutations as the cause of intra- and inter-population variations. Bachry *et al.* (2020) stated that site-specific can be used as the genetic identity of a population. Genetic variation will increase because offspring receive a unique combination of genes and chromosomes from their parents through gene recombination that occurs through sexual reproduction (Indrawan *et al.* 2007; Soewardi 2007; Irmawati 2016).

Halong and Nania populations representing mixed vegetation and monospecific vegetation seagrass habitats each had four haplotypes from four samples with high haplotype (Hd) diversity of 1,000 and nucleotide diversity ( $\pi$ ) of 0.004 and 0.002. While in Poka and Tanjung Tiram, each had one haplotype from three samples with low haplotype and nucleotide diversity of 0.000 (Table 2). Nucleotide diversity is the average number of nucleotide differences per site between two randomly selected individuals from a population (Hughes *et al.* 2008). According to Hughes *et al.* (2008) and Mukhopadhyay and Bhattacharjee (2016), the level of genetic diversity within a population can affect survival, productivity, growth, reproduction, population stability, as well as inter-specific interactions within communities, and processes at the ecosystem level.

Inter-population variation is greater than intra-population with values of 78.81% and 21.19%, respectively (Table 3). Madduppa *et al.* (2019) also found inter-population genetic variation (62.04%) which was higher than intra-population (37%) between the Northern and Southern regions of the Seribu Islands, Jakarta, Indonesia, where the diversity between these areas was thought to be influenced by high fishing pressures and anthropogenic activities such as pollution. According to Mukhopadhyay and Bhattacharjee (2016), genetic variation not only affects individual or population fitness, but also contributes to population fitness and successful recovery.

Each observation station that represents a different seagrass habitat certainly has different environmental characteristics. As reported by Latuconsina *et al.* (2020a), that Poka and Nania are characterised by monospecific seagrass

habitats, each of which is fed by a river, so that they have similar characteristics in the form of typical turbidity values, dissolved oxygen, chlorophyll-a, and high temperatures. While Tanjung Tiram has high levels of phosphate and nitrate, and together with Halong station has high salinity and pH characteristics, this is possible because there is no river influence which can cause pH and salinity fluctuations in Tanjung Tiram and Halong. Differences in environmental conditions can affect the genetic variation of *S. canaliculatus* populations between observation stations, which in turn has the potential to affect its phenotype. According to Li *et al.* (1993) and Mukhopadhyay and Bhattacharjee (2016), environmental conditions, genetic factors, and genetic interactions with the environment can affect phenotypic differences.

Latuconsina *et al.* (2020a) found higher levels of turbidity in seagrass habitats of monospecific vegetation (Poka and Nania) compared to mixed vegetation (Tanjung Tiram and Halong). This phenomenon is supported by Noya *et al.* (2016b) who estimated that the highest cohesive sediment concentrations occurred in the Poka to Nania areas, where river mouths are the main source of sediment material carrier, with sediment transport rates in IAB waters reaching the range of 1.75 cm–10.01 cm or about 39.9 mm/day. Irawan and Nganro (2016) reported that high sedimentation in IAB waters had threatened the existence of seagrass habitats. This phenomenon is thought to affect the adaptability of *S. canaliculatus* populations to each seagrass habitat with different turbidity and sedimentation pressure, which of course has the potential to affect changes and genetic variation between *S. canaliculatus* populations between seagrass habitats.

The genetic differentiation of intra- and inter-population *S. canaliculatus* populations in seagrass habitats in IAB waters is thought to be influenced by tidal currents that support gene flow through individual migration and distribution of pelagic larvae. The current direction pattern that occurs in IAB waters, according to Fadli *et al.* (2014), is strongly influenced by the tides, where during the tidal phase towards the lowest ebb, the water flows out of the IAB waters, on the other hand at low tide towards the highest tide the water flows into IAB waters. According to Swain *et al.* (2004), genetic differences between groups are an indication of genetic differentiation that is influenced by two factors:

1. Gene flow, namely changes in allele frequencies resulting from the movement of gametes between individuals or groups that can reduce or prevent differentiation.
2. Gene drift refers to random fluctuations in frequency. Alleles arise from the gametes sampling in a finite population and can promote genetic differentiation.

5 The genetic diversity of the high population of *S. canaliculatus* between seagrass habitats in IAB waters will be very vulnerable if one of the seagrass habitats is damaged or even disappears due to other anthropogenic activity, including the high exploitation of *S. canaliculatus*. According to Martinez *et al.* (2018), life history

and habitat characteristics play a role in shaping patterns of genetic diversity in fish, so they should be considered in prioritising species for conservation efforts.

Cluster 1 shows the population of Nania (H1, H2, H3 and H4), Cluster 2 shows the population of Halong (H7, H8, H9 and H10), and Cluster 3 consists of the population of Poka (H5) and Tanjung Tiram (H6). The highest variation of nucleotide sites was found at Halong and Nania stations, each of which represented different seagrass habitats, namely mixed vegetation and monospecific vegetation (Fig. 3). Bramandito *et al.* (2018) reported nine haplotypes from several populations of *S. canaliculatus* from the Seribu Islands, Jakarta, Indonesia. Bachry *et al.* (2019) reported six haplotypes in the aquatic animal population of the abalone species *H. squamata* from the southern waters of Java and Bali. The number of haplotypes and the diversity of haplotypes can affect the genetic diversity of a population (Akbar *et al.* 2014).

Ambon Island has a unique geological history, and consists of two different islands, namely Leihitu in the north, which is a volcanic island, and Leitimur in the south which was formed due to tectonic activity. These two distinct islands are connected by a narrow isthmus (Honthaas *et al.* 1999; Pownall *et al.* 2013). This means that geologically, Inner Ambon Bay (IAB) was once united with Baguala Bay in the eastern part of Ambon Island, which in geological history then separated after the merging of these two islands with the existence of a narrow isthmus. This phenomenon might explain that the Nania station (which is in the position of the narrow isthmus that unites Leihitu and Leitimur) has a specific nucleotide site because it is more isolated with a longer genetic distance compared to the other three observation stations (Tanjung Tiram, Halong and Poka).

Madduppa *et al.* (2019) obtain a close genetic distance (0.00–0.06) in the population of *S. canaliculatus* between stations representing the Southern and Northern parts of the Seribu Islands cluster of Jakarta, Indonesia. Sahabuddin *et al.* (2019) found that the genetic variation of the population of *S. canaliculatus* is strongly influenced by the physical distance of a water area. According to Nei (1987), the value of genetic distance is a value scale that describes the similarity of base sequences in the CO1 gene fragment, the smaller the value, the closer the kinship relationship between the two populations. Irmawati (2016) stated that fish individuals that have close genetic distance or high DNA sequence homologs can be categorised as one population.

The cause of the difference in genetic distance values between populations of *S. canaliculatus* is strongly suspected to be caused by the migration of individual fish supported by different current patterns in the IAB waters. Nurfitri and Putri (2019) simulation to look at current patterns in Ambon Bay shows that surface water masses coming and going from and to the IAB are through the narrow west side (Tanjung Tiram) of the inlet/canal (sill). According to Saputra and Lekalette (2016), the pattern of current directions in IAB waters follows a low tide period at a depth of 5 m which moves from the east to the middle of the bay then southwest towards OAB but most are blocked when crossing the threshold. While the flow of water from the OAB during high tide does not originate from the sill towards the IAB, when it passes through the sill, the current is divided into two different

directions: (a) The mass of water entering through the west side (Tanjung Tiram) heading to the northwest (Poka), and at low tide the current pattern from the Poka moves towards the next Poka towards the OAB past the Sill, and (b) on the other water masses to the southeast (Halong), then moves to the northeast towards the centre of the IAB.

This phenomenon can answer the absence of specific nucleotide sites between Tanjung Tiram and Poka stations because the current pattern is thought to help distribute pelagic larvae of the *S. canaliculatus* population to connect to each other at the two stations. In contrast, the Poka and Tanjung Tiram stations with the Halong stations are not connected to each other because there is no current that distributes the pelagic larvae of *S. canaliculatus* to be connected between the three stations. This phenomenon can answer the many specific nucleotide sites in Halong station.

Eddy events often occur with different patterns between the tides and ebbs of sea water, as well as between the rainy and dry seasons in IAB waters. As Noya et al. (2016) and Noya et al. (2019) reported that counterclockwise vertical eddies are dominant in the low tide phase, and in the high tide phase there are clockwise vertical eddies. Water discharge from the estuary that flows into the IAB is thought to be the main factor influencing circulation and eddy patterns in the IAB during the rainy season. Meanwhile, the configuration of topography and tidal parameters during the dry season is thought to be the main factor causing eddies in the IAB. In addition, the dominant anti-clockwise horizontal eddy occurs in the middle of the bay and the eastern part of the IAB, while the dominant clockwise horizontal eddy occurs in the northern part of the IAB. The eddy phenomenon is thought to be one of the physical barriers to the spatial distribution of *S. canaliculatus* pelagic larvae among seagrass habitats, for example between Tanjung Tiram and Poka and Halong, as well as between Nania and other observation stations (Tanjung Tiram, Halong and Poka). There is a relatively large difference in genetic distance between *S. canaliculatus* populations at Nania station and other stations (Table 4), presumably due to the Eddy phenomenon that occurs at several central points of IAB during high and low tide periods as a physical barrier as reported by Noya et al. (2016a), and Salamena et al. (2021). This phenomenon can also explain the discovery of specific genetic nucleotide sites for *S. canaliculatus* at Halong and Nania stations.

Putra and Pratomo (2019) analysed the direction and speed of currents in the waters of Ambon Bay, at high tide, the current speed tends to be small, namely 0.01 m/s and the current direction only rotates in IAB waters, while at the lowest tide, the current speed range is 0.015 m/s–0.030 m/s with the dominant direction towards OAB waters. According to Fadli et al. (2014), movement and current patterns play an important role in changing water masses in Ambon Bay. This phenomenon is thought to play an important role in supporting the distribution of pelagic larvae in different seagrass habitats according to current patterns in IAB waters.

The presence of genetic variation of the *S. canaliculatus* population at each observation station (between seagrass habitats) in IAB waters shows the role of tidal currents as a supporter of the distribution of pelagic larvae in the direction of the current pattern, it also becomes a barrier to the distribution of pelagic larvae that are not in the directions of the current pattern although the distance between adjacent seagrass habitats such as between Tanjung Tiram and Halong stations is only one nautical mile, but separated by a threshold between the OAB, so that the current becomes a barrier to the distribution of larvae and adults of *S. canaliculatus* between adjacent habitats. Rabbitfish (*Siganus* spp.), according to Fisher *et al.* (2005), showed wide dispersal potential supported by ocean currents, with strong swimming ability at speeds of 34.2 cm/s–87.1 cm/s (mean  $67.1 \pm 8.9$ ) during the pelagic larval stage (29.5 mm TL). This phenomenon will certainly support spatial distribution. *S. canaliculatus* in IAB waters are supported by tidal currents. As (Hsu & Gwo 2017) found ocean currents that have a positive contribution in efforts to recover the population of *S. fuscescens* from extreme temperature stress.

Grant and Bowen (1998), stated that marine fish generally show low levels of genetic differentiation between geographic areas due to the potential for higher dispersal during the pelagic egg and larval stages or during the adult phase coupled with the absence of physical barriers to movement between adjacent sites. According to (Lin & Liu 2008), the close genetic distance is also caused by currents, the high dispersal ability of biota larvae, and the availability of supportive habitats (Huyghe & Kochzius 2018). Madduppa *et al.* (2019) revealed that genetic distance is an assessment of kinship relationships between populations from several locations, where the lower the value of genetic distance obtained indicates a very close relationship between locations, and the higher the value indicates a fairly distant relationship between locations.

Based on the phylogenetic tree, the *Siganus* spp. and Outgroup *Lutjanus russellii* depicts two main clusters. Cluster 1 shows two main subclusters consisting of subcluster (1-a) of intrapopulation *S. canaliculatus* from IAB and subcluster (1-b) from genbank (*S. canaliculatus*, *S. fuscescens*, *S. sutor*) originating from other geographical areas (Fig. 3).

Interestingly, the intrapopulation at Halong is phylogenetically separated among the four sequences of *S. canaliculatus* (Fig. 3), where Halong-2 and Halong-3 are more closely related to the Nania population, while Halong-1 and Halong-4 are more closely related to the Tanjung Tiram population. It is suspected that the geographical condition of Halong station is more open and faces an inlet that separates IAB waters from OAB, so that the opportunity for gene migration from inside and outside the IAB waters is greater through crossbreeding. According to Soewardi (2007), genetic differences between populations are supported by gene flow between sub-populations and genetic drift from certain sub-populations.

The accuracy of the kinship test based on the CO1 gene sequence can be relied on. Tree clades represent relationships between units by tracing the paths of descent from ancestors. Li and Graur (2000) revealed that the branch

length indicates the number of evolutionary changes that occurred between the two clusters. The diversity of CO1 gene sequences has the potential to identify species and DNA barcodes (Hebert *et al.* 2003; Hebert & Gregory 2005).

The results of this study confirm that the *S. canaliculatus* intraspecies is phylogenetically separate from other subspecies of *S. canaliculatus*, although the results are still in the same cluster. Meanwhile, *S. canaliculatus* intraspecies is still a sub-species of the genus *Siganus*. Our suggestion is to collect more molecular data from this species distribution to establish a stronger status for protected taxa.

## CONCLUSION

The pattern and direction of tidal currents with the eddy phenomenon as a link or barrier to spatial distribution and connectivity of *S. canaliculatus* larvae between seagrass habitats, as well as the presence of different anthropogenic pressures in each seagrass habitat, are thought to influence the genetic characteristics (genetic diversity, genetic variation, genetic differentiation and genetic distance) of *S. canaliculatus* populations in IAB waters.

Preliminary research using mitochondrial CO1 sequences can at least explain that the characteristics of seagrass habitat and its geographical location in narrow bay areas such as in IAB can affect the diversity, variation, differentiation and genetic distance of *S. canaliculatus* populations. Thus, further in-depth and comprehensive research is still needed regarding population genetics and inter-population connectivity of *S. canaliculatus* in the waters of IAB and outside IAB, with a more representative number of samples using microsatellites, so that it is expected to complement the results of the preliminary research that has been done.

The results of this research can provide information about the urgency of habitat-based fisheries management of *S. canaliculatus* through the development of conservation areas based on the characteristics of seagrass habitats and the biological characteristics of *S. canaliculatus* populations, including their genetic characteristics, to support sustainable utilisation in IAB waters.

## ACKNOWLEDGEMENTS

Our gratitude goes to the Universitas Islam Malang for the funding assistance to accelerate the completion of Doctoral studies at the Program Study of Aquatic Resources Management, Graduate School, IPB University, and the Educational Fund Management Institute, Ministry of Finance of the Republic of Indonesia (LPDP) for the assistance of 2019 Fiscal Year Research Dissertation Research Scholarship No. PRJ-32/LPDP.4/2019.



## AUTHORS' CONTRIBUTIONS

Husain Latuconsina: Conception and design of study, collection and preservation of fish samples in the field, analysis and interpretation of data, drafting the manuscript, revising the manuscript for significant intellectual content.

Nurlisa A. Butet: Conception and design of study, revising the manuscript.

Ridwan Affandi: Conception and design of study, revising the manuscript.

M. Mukhlis Kamal: Conception and design of study, revising the manuscript.

Syamsul Bachry: Analysis of data, revising the manuscript.

Agus A. Hakim: Genetic test fish samples in the laboratory (isolation, purification, and amplification).

## REFERENCES

- Akbar N, Zamani N P and Madduppa H H. (2014). Genetic diversity of yellowfin tuna (*Thunnus albacares*) from two populations in the Moluccas Sea, Indonesia. *Depik* 3(1): 65–73. <https://doi.org/10.13170/depik.3.1.1304>
- Allen G R and Erdmann M V. (2012). *Reef fishes of the East Indies*. Vol. I–III. Perth, Australia: Tropical Reef Research, 1292 pp.
- Ambo-Rappe R, Nessa M N, Latuconsina H and Lajus D L. (2013). Relationship between the tropical seagrass bed characteristics and the structure of the associated fish community. *Open Journal of Ecology* 3(5): 331–342. <https://doi.org/10.4236/oje.2013.35038>
- Bachry S, Solihin D D, Rudhy G, Kadarwan S, and Butet N A. (2019). Genetic diversity of the *Haliotis diversificolor squamata* from Southern Coastal Java (Banten, Pangandaran and Alas Purwo) and Bali Based on Mitochondrial CO1 sequences. *Tropical Life Sciences Research* 30(3): 83–93. <https://doi.org/10.21315/tlsr2019.30.3.6>
- \_\_\_\_\_. (2020). Filogeni populasi *Haliotis squamata* Reeve, 1846 dari Pantai Selatan Pulau Jawa dan Bali berdasarkan sekuen cytochrome B DNA mitokondria (Phylogenetic of *Haliotis squamata* Reeve, 1846 population from the Southern Coast of Java and Bali Island in Indonesia based on cytochrome B mitochondrial DNA sequence). *Jurnal Ilmu dan Teknologi Kelautan Tropis* 12(2): 585–595. <https://doi.org/10.29244/jitkt.v12i2.30691>
- Bandelt H J, Forster P and Röhl A. (1999). Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* 16(1): 37–48. <https://doi.org/10.1093/oxfordjournals.molbev.a026036>
- Basit A, Putri M R and Tatipatta W M. (2012). Estimation of seasonal vertically integrated primary productivity in Ambon Bay using the depth-resolved, time-integrated production model. *Marine Research in Indonesia* 37(1): 47–56.
- Bramandito A, Subhan B, Prartono T, Anggraini N P, Januar H I and Madduppa H H. (2018). Genetic diversity and population structure of *Siganus fuscescens* across urban reefs of Seribu Islands, Northern of Jakarta, Indonesia. *Biodiversitas Journal of Biological Diversity* 10(6): 1993–2002. <https://doi.org/10.13057/biodiv/d190603>
- Evans S M, Dawson M, Dya J, Frid C L J, Gill M E, Pattiasina L A and Porter J. (1995). Domestic waste and TBT pollution in coastal areas of Ambon Island (Eastern Indonesia). *Marine Pollution Bulletin* 30(2): 109–115.

- Excoffier L, Smouse P E and Quattro J M. (1992). Analysis of molecular variance inferred from metric distances among DNA haplotypes: Application to human mitochondrial DNA restriction data. *Genetics* 131: 479–491. <https://doi.org/10.1093/genetics/131.2.479>
- Fadli M, Radjawane I M and Susanna. (2014). Pemodelan hidrodinamika di perairan Teluk Ambon. In A S Atmadipoera, I Jaya, S M Natsir, N Hendriarti, B Herunadi, M P Patria, R Zuraida, K T Dewi, W Pranowo, T Prartono, et al. (eds.). *Prosiding Pertemuan Ilmiah Nasional Tahunan X ISOI 2012*. Jakarta: Ikatan Sarjana Oseanologi Indonesia (ISOI), 6–19.
- Fisher R, Leis J M, Clark D L and Wilson S K. (2005). Critical swimming speeds of late-stage coral reef fish larvae: Variation within species, among species and between locations. *Marine Biology* 147: 1201–1212. <https://doi.org/10.1007/s00227-005-0001-x>
- Gemilang W A, Rahmawan G A and Wishwa U J. (2017). Kualitas perairan Teluk Ambon dalam berdasarkan parameter fisika dan kimia pada musim peralihan I (Inner Ambon Bay water quality based physical and chemical parameters in transition season I). *EnviroSciencieae* 13(1): 79–90. <https://doi.org/10.20527/es.v13i1.3518>
- Grant W S and Bowen B W. (1998). Shallow population histories in deep evolutionary lineages of marine fishes: Insights from sardines and anchovies and lessons for conservation. *Journal of Heredity* 89(5): 415–426. <https://doi.org/10.1093/jhered/89.5.415>
- Hajibabaei M, Smith M A, Janzen D H, Rodriguez J J, Whitfield J B and Hebert P D N. (2006). A minimalist barcode can identify a specimen whose DNA is degraded. *Molecular Ecology* 6: 959–964. <https://doi.org/10.1111/j.1471-8286.2006.01470.x>
- Hebert P D N and Gregory T R. (2005). The promise of DNA barcoding for taxonomy. *Systematic Biology* 54(5): 852–859. <https://doi.org/10.1080/10635150500354886>
- Hebert P D N, Ratnasingham S and de Waard J R. (2003). Barcoding animal life: Cytochrome c oxidase subunit 1 divergences among closely related species. *Proceedings: Biological Sciences* 270: S96–S99.
- Honthaas C, Maury R C, Priadi B, Bellon H and Cotton J. (1999). The Plio–Quaternary Ambon arc, Eastern Indonesia. *Tectonophysics* 301(3–4): 261–281. [https://doi.org/10.1016/S0040-1951\(98\)00227-3](https://doi.org/10.1016/S0040-1951(98)00227-3)
- Hsu T H and Gwo J C. (2017). Fine-scale genetic structure of rabbitfish, *Siganus fuscescens*, in Penghu Archipelago following a mass mortality event caused by extreme cold winter weather. *Genes and Genomics* 39: 645–652. <https://doi.org/10.1007/s13258-017-0530-y>
- Hughes A R, Inouye B D, Johnson M T J, Underwood N and Vellend M. (2008). Ecological consequences of genetic diversity. *Ecology Letters* 11: 609–623. <https://doi.org/10.1111/j.1461-0248.2008.01179.x>
- Huyghe F and Kochzius M. (2018). Sea surface currents and geographic isolation shape the genetic population structure of a coral reef fish in the Indian Ocean. *PLoS ONE* 13(3): e0193825. <https://doi.org/10.1371/journal.pone.0193825>
- Ikhsani I Y, Abdul M S and Lekalette J D. (2016). Distribusi fosfat dan nitrat di Teluk Ambon bagian dalam pada monsun barat dan timur (Phosphate and nitrate distribution in inner Ambon Bay during northwest and southeast monsoon). *Widyariset* 2(2): 86–95. <https://doi.org/10.14203/widyariset.2.2.2016.86-95> [Indonesia]
- Indrawan M, Primack R B and Supriatna J. (2007). *Biologi Konservasi (Conservation biology)*. Jakarta: Yayasan Obor Indonesia.

- Irawan A and Nganro M N. (2016). Sebaran lamun di Teluk Ambon Dalam (Distribution of Seagrasses in Inner Ambon Bay). *Jurnal Ilmu dan Teknologi Kelautan Tropis* 8(1): 99–114. <https://doi.org/10.29244/jitkt.v8i1.12499>. [Indonesia]
- Imawati. (2016). *Genetika populasi ikan (Fish population genetics)*. Yogyakarta: Andi Offset.
- Kumar S, Stecher G, Li M, Knyaz C and Tamura K. (2018). MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35(6): 1547–1549. <https://doi.org/10.1093/molbev/msy096>
- Latuconsina H, Affandi R, Kamal M M and Butet N A. (2020a). Distribusi spasial ikan baronang *Siganus canaliculatus* Park, 1797 pada habitat padang lamun berbeda di Teluk Ambon Dalam (Spatial distribution of white-spotted rabbitfish *Siganus canaliculatus* (Park, 1797) in different seagrass beds habitat in Inner Ambon Bay). *Jurnal Ilmu Dan Teknologi Kelautan Tropis* 12(1): 89–106. <https://doi.org/10.29244/jitkt.v12i1.27908>
- \_\_\_\_\_. (2020b). On the assessment of white-spotted rabbitfish (*Siganus canaliculatus* Park, 1797) stock in the Inner Ambon Bay, Indonesia. *AACL Bioflux* 13(4): 1827–1835.
- Latuconsina H, Ambo-Rappe R and Burhanuddin A I. (2023). *Ichthyofauna of tropical seagrass meadows: Biodiversity, threats, and their management*. Yogyakarta. Gadjah Mada University Press.
- Latuconsina H, Ambo-Rappe R and Nessa M N. (2013). Association of rabbit fish (*Siganus canaliculatus* Park, 1797) in the seagrass ecosystems in the Inner Ambon Bay. In C P H Simanjuntak (ed.). *Proceedings of the National Fish Seminar VII*. Indonesian Ichthyology Society, 123–137 [Indonesian].
- Latuconsina H, Kamal, M M, Affandi R, and Butet N A. (2022). Growth and reproductive biology of white-spotted rabbitfish (*Siganus canaliculatus*) on different seagrass habitats in Inner Ambon Bay, Indonesia. *Biodiversitas Journal of Biological Diversity* 23(1): 273–285. <https://doi.org/10.13057/biodiv/d230133>
- Li S, Cai W and Zhou B. (1993). Variation in morphology and biochemical genetic markers among populations of blunt snout bream (*Megalobrama amblycephala*). *Aquaculture* 111(1–4): 117–127.
- Li W H and Graur D. (2000). *Fundamentals of molecular evolution*, 2nd ed. Sunderland: Sinauer Associates Inc,
- Lin T and Liu L. (2008). Low levels of genetic differentiation among populations of the coral-inhabiting snail *Coralliophila violacea* (Gastropoda: Coralliophilidae) in regions of the Kuroshio and South China Sea. *Zoological Studies Taipei* 47(1): 17–24.
- Madduppa H H, Margaretha M K, Bramandito A, Prartono T, Subhan B, Arafat D and Anggraini N P. (2019). Short communication: Intraspecific genetic diversity and population subdivision of rabbitfish (Siganidae: *Siganus canaliculatus*) in urbanized reefs of Jakarta Bay, Indonesia. *Biodiversitas Journal of Biological Diversity* 20(10): 2897–2902. <https://doi.org/10.13057/biodiv/d201017>
- Mahmudi M, Lusiana E D, Herawati E Y and Seriholo L G. (2020). Environmental factors and seasonal effect on the potential harmful algae presence at Ambon Bay, Indonesia. *Biodiversitas Journal of Biological Diversity* 21(7): 3101–3107.
- Manik N. (1998). Estimation of growth parameters and mortality of Rabbit fish (*Siganus canaliculatus*) in Inner Ambon Bay. *Jurnal Perairan Maluku dan Sekitarnya* 12: 55–63. [Indonesia]
- Manullang C Y, Barends W, Polnaya D, Soamole A and Rehalat I. (2021). Marine litter and grading of the coastal areas of Ambon Bay, Indonesia. *Jurnal Omni-Akuatika* 17(2): 70–77.

- Markert J A, Champlin D M, Gutjahr-Gobell R, Grear J S, Kuhn A, McGreevy T J and Nacci D E. (2010). Population genetic diversity and fitness in multiple environments. *BMC Ecology and Evolution* 10: 205. <https://doi.org/10.1186/1471-2148-10-205>
- Martinez A S, Willoughby J R and Christie M R. (2018). Genetic diversity in fishes is influenced by habitat type and life-history variation. *Ecology and Evolution* 2018: 1–10. <https://doi.org/10.1002/ece3.4661>
- Mukhopadhyay T and Bhattacharjee S. (2016). Genetic diversity: Importance and measurements. In A H Mir and N A Bhat (eds.). *Conserving biological diversity: A multiscaled approach*. New Delhi, India: Research India Publications, 251–295.
- Nei M. (1987). *Molecular evolutionary genetics*. New York: Columbia University Press.
- Noya Y A, Kalay D E, Purba M, Koropitan A F and Prartono T. (2019). Modelling baroclinic circulation and particle tracking in Inner Ambon Bay. *IOP Conference Series: Earth and Environmental Science* 339: 012021. <https://doi.org/10.1088/1755-1315/339/1/012021>
- Noya Y A, Purba M, Koropitan A F and Prartono P. (2016a). Modeling the barotropic circulation on Inner Ambon Bay. *International Journal of Oceans and Oceanography* 10(2): 265–286.
- \_\_\_\_\_. (2016b). Cohesive sediment transport modeling on Inner Ambon Bay. *Journal of Tropical Marine Science and Technology* 8(2): 671–687. <https://doi.org/10.29244/jitkt.v8i2.15834>
- Nurfitri S and Putri M R. (2019). Study on water mass exchange at Ambon Bay using trajectory model: Circulation of one tidal cycle. *IOP Conference Series: Journal of Physics: Conference Series* 1127(2019): 012039. <https://doi.org/10.1088/1742-6596/1127/1/012039>
- Pello F S, Adiwilaga E M, Huliselan N V and Damar A. (2014). Pengaruh musim terhadap beban masukan nutrien di Teluk Ambon Dalam (Effect of seasonal on nutrient load input the Inner Ambon Bay). *Jurnal Bumi Lestari* 14(1): 63–73.
- Pownall J M, Hall R and Watkinson I M. (2013). Extreme extension across Seram and Ambon, eastern Indonesia: Evidence for Banda slab rollback. *Solid Earth* 4: 277–314. <https://doi.org/10.5194/se-4-277-2013>
- Putra H S and Pratomo D G. (2019). Analysis of sediment flow and transport using 3-dimensional hydrodynamic modeling (Case study: Ambon Bay, Ambon City, Maluku). *Journal of Eng ITS* 8(2): 124–129. [Indonesian]
- Rahmawan G A, Gemilang W A, Wisha U J, Shiauddin R and Ondara K. (2019). Estimation of sediment distribution based on bathymetry alteration (204-2016) in the Inner Ambon Bay of Ambon, Maluku, Indonesia. *Jurnal Segara* 15(2): 67–78. <https://doi.org/10.15578/segara.v15i2.6956>
- Rozas, J. (2009). DNA Sequence Polymorphism Analysis Using DnaSP. In D Posada. (Ed.), *Bioinformatics for DNA sequence analysis*. New Jersey: Humana Press, 337–350. [https://doi.org/10.1007/978-1-59745-251-9\\_17](https://doi.org/10.1007/978-1-59745-251-9_17)
- Sahabuddin, Burhanuddin A I, Tuwo A and Malina A C. (2019). Genetic variation of rabbit fish (*Siganus canaliculatus* Park, 1797) in the waters of Bone Bay and Makassar Strait. *International Journal of Scientific and Technology Research* 6(6): 171–181. <https://doi.org/10.32628/IJSRST196620>
- Salamena G G, Whinney J C, Heron S F and Ridd P V. (2021). Internal tidal waves and deep-water renewal in a tropical fjord: Lessons from Ambon Bay, eastern Indonesia. *Estuarine, Coastal and Shelf Science* 253(2021): 107291. <https://doi.org/10.1016/j.ecss.2021.107291>
- Sambrook J, Fritsch E R and Maniatis T. (1989). *Molecular cloning: A laboratory manual*, 2nd ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.

- Saputra F R T and Lekalette J D. (2016). Water mass dynamics in Ambon Bay. *Widyariset* 2(2): 143–152. <https://doi.org/10.14203/widyariset.2.2.2016.143-152>
- Selano D A J, Adiwilaga E M, Dahuri R, Muchsin I and Effendi H. (2009). Sebaran spasial luasan area tercemar dan analisis beban pencemar bahan organik pada perairan Teluk Ambon Dalam (Spatial distribution of pollution areas and analysis of pollution load of organic matter in Inner Ambon Bay waters). *Torani Journal of Fisheries and Marine Science* 19(2): 96–106.
- Soewardi. (2007). *Pengelolaan keragaman genetik sumberdaya perikanan dan kelautan*. Indonesia: Department of Aquatic Resources Management, Bogor Agricultural Institute.
- Swain D P, Hutchings J A and Foote C. (2004). Environmental and genetic influences on stock identification characters. In S X Cadrin, L A Kerr and S Mariani (eds.). *Stock identification methods applications in fishery science*. San Diego, CA: Elsevier Academic Press, 45–85.
- Toro M and Caballero A. (2005). Characterization and conservation of genetic diversity in subdivided populations. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* 360: 1367–1378. <https://doi.org/10.1098/rstb.2005.1680>
- Vogelstein B and Gillespie D. (1979). Preparative and analytical purification of DNA from agarose. *Proceedings of the National Academy of Sciences of the United States of America* 76: 615.
- Ward R D, Costa F O, Holmes B H and Steinke D. (2008). DNA barcoding of shared fish species from the North Atlantic and Australasia: Minimal divergence for most taxa, but *Zeus faber* and *Lepidopus caudatus* each probably constitute two species. *Aquatic Biology* 3(1): 71–78. <https://doi.org/10.3354/ab00068>
- Ward R D, Zemlak T S, Innes B H, Last P R and Hebert P D N. (2005). DNA barcoding Australia's fish species. *Philosophical Transactions of the Royal Society of London Series B – Biological Sciences* 360: 1847–1857. <https://doi.org/10.1098/rstb.2005.1716>
- Woodland D J (2001). Siganidae, Rabbitfishes (spinefoots). In K E Carpenter and V Niem (eds.), *FAO species identification guide for fishery purposes, The living marine resources of the Western Central Pacific*, Vol. 6, Bony fishes part 4 (Labridae to Latimeriidae), estuarine crocodiles. Rome: FAO, 3627–3650.

# Genetic Diversity of White-spotted Rabbitfish (*Siganus canaliculatus*) on Different Seagrass Habitats in Inner Ambon Bay, Indonesia Based on Mitochondrial CO1 Sequences

## ORIGINALITY REPORT

7%

SIMILARITY INDEX

7%

INTERNET SOURCES

5%

PUBLICATIONS

3%

STUDENT PAPERS

## PRIMARY SOURCES

1

[www.tlsr.usm.my](http://www.tlsr.usm.my)

Internet Source

2%

2

[www.bioflux.com.ro](http://www.bioflux.com.ro)

Internet Source

2%

3

[mail.smujo.id](mailto:mail.smujo.id)

Internet Source

1%

4

[biodiversitas.mipa.uns.ac.id](http://biodiversitas.mipa.uns.ac.id)

Internet Source

1%

5

S N N Putri, G R Maharani, M Farhan, D F Lestari et al. "Population connectivity and genetic diversity population connectivity of Rabbitfish (*Siganus canaliculatus*) among Bangladesh, China and Indonesia", IOP Conference Series: Earth and Environmental Science, 2022

Publication

1%

6

[www.scielo.sa.cr](http://www.scielo.sa.cr)

Internet Source

1%

---

Exclude quotes On

Exclude matches < 1%

Exclude bibliography On